Passive eDNA collection enhances aquatic biodiversity analysis

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Motivation

Environmental DNA (eDNA) Metabarcoding

- Novel method for assessing biodiversity wherein samples are taken from the environment via water, sediment or air from which DNA is extracted, and then amplified using general or universal primers in polymerase chain reaction and sequenced using next-generation sequencing to generate thousands to millions of reads.
- Metabarcoding removes the need for multiple taxonomic experts by automatically matching the DNA samples to a taxonomic identity from an existing database
- Because the identity matching is automated, the limitations come purely from how many samples can be metabarcoding

Motivation

Room for Improvement?

- Main objective \rightarrow more samples
- Increase the amount of samples that one can collect by switching from active filtering to passive filtering

Active Filtering

- Collect water samples (1 L 20 L, depends on the environment) and actively pump the water through membranes to collect eDNA samples
- Extremely time and energy intensive, requiring specialized equipment

Design and Implementation

Their Approach

- They present the alternative of switching from the active pump system to a passive membrane collection system
- Using two membranes:
 - **positively charged nylon** to catch eDNA particles by charge attraction
 - non-charged cellulose ester to catch eDNA particles by entrapment
- They attached these membranes to an oyster aquaculture frame with mesh pockets and submerged underwater



Evaluation

Varying Climate

- They tested their system in two different climates
 - Tropical Waters in the Ashmore Reef
 - Temperate Waters around Daw Island

Varying Collection Time

 They tested how varying the particle collection time affected the overall taxa identification

Compared Against Active Filtering in all Cases

 In all tests at both locations they compared against active filtering (using 9 L of water) as their ground truth



Sample Post Processing

Standard Methods Used for all eDNA Sequencing

- One-step quantitative polymerase chain reaction (qPCR) were performed with each sample and a universal primer
 - Primer is picked based on the fact that they're targeting fish taxa
- PCR outputs also include controls
 - **Positive Control:** DNA sample of a fish that should not be in the environment that they collected, but should be identified with 100% accuracy
 - All identified the known fish with 100% accuracy
 - Minimum reads in the positive control was 36, therefore a conservative cutoff for their application was 40 reads
 - **Negative Control:** use deionized water instead of DNA sample
 - No sample produced more than 5 reads for any species
- They compare these outputs against a database of known fish (different for each region)
 - < 80% match: sample discarded
 - 80 % < match < 90 % : family
 - 90 % < match < 97 % : genus
 - > 97 % : species

Supplementary Data and Results - Ashmore

Table 1 Taxa dete	cted at Ashmore Reef.										Table 1 (continue	d)												
Family name	T	Deer								A stive filtration	Family name	Taxon name	Passi	ve filtra	ation						Active filtration			
ranny name	Taxon name	Charged Non-charged								- Active hitration			Charg	Charged				Non-charged			_			
		4 h	9 L	12.6	24 h	- 46	0 h	12 h	24 h	-			4 h	8 h	12 h	24 h	4 h	8 h	12 h	24 h				Pa Ch 4 I
Acanthuridae	Acanthurus blochii Acanthurus lineatus	4 1	01	12 11	24 11	41	on	B	B	B	Labridae	Anampses sp. Chlorurus sp. Chaeradan schoenleinii									A A			
	Acanthurus triostegus Acanthurus xanthopterus		B	B B	B	B B	B	В	B	B B		Cirrhilabrus exquisitus Cymolutes praetextatus	в	В			В	В	В	В	A B			
	Ctenochaetus binotatus Ctenochaetus striatus Naso annulatus Naso brachycentron		В		В		В	В Р		B A		Epibulus sp. Gomphosus varius Halichoeres hortulanus Halichoeres melanurus		Р				Ρ			A			
	Naso unicornis Zebrasoma sp. Zebrasoma scopas									A A A		Halichoeres nebulosus Halichoeres trimaculatus Hipposcarus longiceps		B B	В		В	B B	B B	В	A B B			
Albulidae Apogonidae	Albula glossodonta Apogon sp. Jaydia sp.					В	В	B B B	В	B B B		Labrichthys unilineatus Novaculichthys taeniourus Pseudocheilinus evanidus		В	B B		В			B P	B	Table 1 (continued)	
Atherinidae	Arripis georgianus Hypoatherina temminckii				В	B	В		В	В		Pseudocheilinus hexataenia Scarus dimidiatus								В	A	Family name	Taxon name	Pa
Balistidae	Balistapus undulatus Balistidae—unknown 1 Melichthys niger					P B	B B	P B	P B	B B		Scarus niger Scarus psittacus Scarus sp.									A A A			Ch 41
	Rhinecanthus aculeatus Rhinecanthus rectangulus Rhinecanthus verrucosus		В		В	В	В	В	B P	B		Stethojulis bandanensis Stethojulis strigiventer Stethojulis sp.							B B	B B B	B B	Pomacentridae	Abudefduf sexfasciatus/vaigiensis Acanthochromis sp. Amblyglyphidodon curacao Chamia atsiaastanlis	
Belonidae	Platybelone argalus Strongylura incisa Tylosurus crocodilus		В	В		В	B B	B B	В	A B B		Stethojulis trilineata Thalassoma amblycephalum Thalassoma lunare					P B	В		в	B B		Chromis atripectorais Chromis lepidolepis Chromis ternatensis	
Blenniidae	Aspidontus taeniatus Atrosalarias holomelas Blenniella periophthalmus Cirripectes sp.				В	В		Р	В	A A B	Lethrinidae	Thalassoma sp. Lethrinus nebulosus Lethrinus obsoletus Lethrinus sp.1					в	B		В	B A A B		Chromis sp. Chromis viridis Chrysiptera glauca Chrysiptera rex	
Carangidae	Salarias fasciatus Caranx ignobilis Carangidae—unknown 1		в	В	В	В		B	B	B B B		Lethrinus sp.2 Lethrinus variegatus Monotaxis arandoculis					0	5		U	A A A		Chrysiptera sp. Dascyllus aruanus Dascyllus reticulatus	
Chaetodontidae	Chaetodon auriga Chaetodon vagabundus Chaetodon sp.						В	Р		B A	Lutjanidae	Aprion virescens Caesio caerulaurea Caesio xanthonotus	В				В	В	B P		B		Dascyllus trimaculatus Dischistodus prosopotaenia Hemiglyphidodon plagiometopon Neopomacentrus sp.	
Cirrhitidae Clupeidae	Paracirrhites forsteri Amblygaster sirm Spratelloides delicatulus	В	В	В	В	В	B B	в	В	A B B		Lutjanus bohar Lutjanus decussatus Lutjanus fulvus		В	В		В	В	B P	В	B A		Plectroglyphidodon lacrymatus Plectroglyphidodon leucozonus Pomacentrus bankanensis	
Congridae	Gnathophis sp.	-							r	A		Lutjanus kasmira Lutjanus sp. 1		В	В		в	В	В	B	B		Pomacentrus replaogenys Pomacentrus pavo	
Fistulariidae	Platax orbicularis Platax teira Fistularia commersonii	P	В	В	В	Р	В	В	В	B A		Lutjanus sp. 2 Pterocaesio sp. Pterocaesio tile	B	В			B	В	В	В	B B A		Pomacentrus sp. 2 Stegastes fasciolatus Stegastes naricans	
Gadidae Gobiidae	Micromesistius sp. Bryaninops sp. Eviota sp. 1			Р						A A	Monacanthidae Mugilidae	Cantherhines dumerilii Monacanthus chinensis Crenimuqil crenilabis	Ρ				Ρ	P P				Pseudochromidae Rhynchobatidae Schindleriidae	Pseudochromis sp. Rhynchobatus sp. Schindleria sp.	
	Eviota sp. 2 Exyrias sp. Gobiodon sp.					В	В	В		B A A		Ellochelon vaigiensis Mulloidichthys vanicolensis Parupeneus barberinus		Ρ			В	В	Ρ	В	B	Scombridae	Auxis sp. Euthynnus sp. Scomberomorus commerson	
	Gobiidae—unknown 1 Gobiidae—unknown 2 Gobiidae—unknown 3						Ρ			A A		Parupeneus chrysopleuron Parupeneus multifasciatus Parupeneus sp.	В	в			Ρ	в		в	AB	Scorpaenidae Serranidae	Scorpaenodes guamensis Aethaloperca rogaa Cephalopholis argus Caphalopholis Japardur	
	Gobiidae—unknown 4 Paragobiodon sp. Vanderhorstia ornatissima									A A A	Muraenidae	Upeneus tragula Echidna nebulosa Gymnothorax buroensis	Ρ					В	в		B A		Epinephelus sp. Plectropomus laevis Pseudogramma polyacanthus	В
Haemulidae Hemiramphidae Holocentridae	Plectorhinchus chaetodonoides Hyporhamphus sp. Myripristis botche	в				B	В	В	B B B	B B B	Myctophidae	Gymnothorax flavimarginatus Gymnothorax sp. Diaphus watasei					В	В	В		A B A	Soleidae Synodontidae	Variola louti Pardachirus pavoninus Trachinocephalus myops	
Kuhliidae	Myripristis murdjan Sargocentron rubrum Kuhlia sp.									A A A	Myliobatidae Platycephalidae	Myctophidae—unknown 1 Aetobatus ocellatus Sungaocia otaitensis	В	В	В	В	В	B	в	В	A B	Tetraodontidae Xiphiidae	Arothron mappa Arothron stellatus Xiphias gladius	
Kyphosidae	Kyphosus sp.					В	В	В	В	В		Plesiops sp.						В	В		В	Taxa at Ashmore Reef detection those detected by passive fit	ted by passive filtration treatment and submersion Itration only and B indicates taxa detected by bot	n time th met

sive filtration

8h 12h 24h

Non-charged

4h 8h

Supplementary Data and Results - Daw

Table 2 Taxa detect	ed at Daw Island.												Table 2 (continued)	(
Family name	Taxon name	Passive f	filtration									Active	Family name	Taxon name Passive filtration											Active
		Charged	Charged Non-charged filtratio												Charged	l					Non-charged				filtration
		4 h	8 h	12 h	24 h	34 h	4 h	8 h	12 h	24 h	34 h				4 h	8 h	12 h	24 h	34 h	4 h	8 h	12 h	24 h	34 h	
Aplodactylidae Arripidae	Aplodactylus sp. Arripis georgianus Arripis sp.	B B	B	B B B	B B	B B	B B B	B B B	B B B	B B B	B B	B B B	Kyphosidae	Girella sp. Kyphosus gladius/ sydneyanus	В	B	B B	BB		B B	B B	B	B B	B B	BB
Berycidae Bothidae	Latropiscis purpurissatus Centroberyx sp. Lophonectes sp.	Ρ	в		в		в		Р			в	Labridae	Scorpis sp. Achoerodus sp. Austrolabrus maculatus	B B	B B	В	В	в	B B	B B	B B	в	В	B B
Callionymidae	Repomucenus calcaratus Pseudocaranx sp	P	в	в	в		P	в	P	P		в		Bodianus sp. Eupetrichthys anaustipes	В	В	В	P B		P B	В	В	В		В
contingitute	Pseudocaranx wrighti Seriola Ialandi		B	B	5	в	B	5	R	^b	в	B		Halichoeres brownfieldi Labridae—	В	В	В	B		В	В	B	В	В	B
Cheilodactylidae	Trachurus sp. Cheilodactylus sp.		в	Б	В	Б	P		Б		Б	В		unknown 1 Notolabrus fucicola		В	P	5		В		5		P	В
	Cheilodactylidae— unknown 1 Nemadactylus	Р	Р				Р	р		P				Notolabrus parilus Ophthalmolepis lineolata		в	В	В		B	2	в	В	B	В
Chironemidae	valenciennesi Chironemus georgianus	Ρ		Ρ			Р	Р			Р		Lamnidae	Pictilabrus laticlavius Carcharodon	В	B P	В	В		В	В	В	В	В	В
Clinidae	Chironemus maculosus Heteroclinus	в	В	В	B				B	в		B	Monacanthidae	carcharias Acanthaluteres sp. Monacanthidae—	В	В	в			В	в	B B	В	B	B
	adelaidae Heteroclinus eckloniae						В					В		unknown 1 Nelusetta ayraudi Scobinichthys	Ρ		Ρ					P B			в
Clupeidae	Clupeidae— unknown 1 Sardinops saaax	B	B	в	в	в	в	B	в	в	в	B	Moridae	granulatus Lotella rhacina Pseudophycis	В										B A
Congridae	Gnathophis longicauda			P								-	Mullidae	barbata Upeneichthys sp. Upeneichthys stotti	B	R	в	B	В	в	R	B	B	В	B
Dinolestidae	Bathytoshia brevicaudata Dinolestes lewini			в	В							A	Myliobatidae Odacidae	Myliobatis australis Heteroscarus	в	Б	BB	B		BB	В	BB	в		B B
Dussumieriidae	Etrumeus jacksoniensis Enoroulio oustrolio	В		В	B		В	B	P	В		В		acroptilus Olisthops cvanomelas	В	В	В			В	В	В	В	В	В
Enoplosidae Gerreidae	Engrauns austrans Enoplosus armatus Parequula melbournensis	Б	В	В	Б			В	Б	B B		B B	Pempheridae	Siphonognathus sp. Parapriacanthus elongatus	B B	B B	В	B B		B B	B B	B B	B B		B B
Hemiramphidae Isonidae	Hyporhamphus melanochir Iso rhothophilus	В		В		В	В	В	В	В	В	B A	Pinguipedidae	Pempheris sp. Parapercis haackei Parapercis ramsayi	B B	B P	B B P	В		В	B B			В	B B

Table 2 (continued)																
Family name	Taxon name	Passive	Passive filtration													
		Charged	1				Non-cha	filtration								
		4 h	8 h	12 h	24 h	34 h	4 h	8 h	12 h	24 h	34 h					
Platycephalidae	Leviprora inops	P														
	Platycephalus grandispinis	В			В			В	В			В				
Pomacentridae	Chromis sp.	P			Р		Р				P					
	Parma microlepis	В	В				В	В	В	В		В				
Scombridae	Scombridae- unknown 1						В					В				
	Scomber sp.	В	В		В		В	В		В		В				
Scorpaenidae	Scorpaenidae-						Р									

Results - Analyzing Mean Values

Mean Taxa Detected Ashmore -

- Charged: 3
- Non-charged: 10
- Active: 42

Daw -

- Charged: 8
- Non-charged: 11
- Active: 17

Mean Taxa Detection Based on Submersion Time Ashmore -

- After 4 Hours: 2
- After 8 Hours: 5

Daw -

• No significant differences between any of the filters, including active

Results - Analyzing Taxa Community



Conclusion

Their results show **promising evidence** that it could be used to properly collect eDNA and **significantly expand the amount of environmental metabarcoding** that can be done and biodiversity that can be analyzed.

The passive solution is

- Inexpensive and scalable
 - Eliminates any need for active / manual collection and filtration
- More appropriate for temperate, but still acceptable for tropical environments
- Easily replicable for more analysis on viability as well as implementation
 - Different membrane materials, understanding physical limitations of membranes, understanding implications of varying environments